See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/303121364

Modulatory effect of fenugreek loaded PLGA Nanoparticles on Lipofuscinogenesis in pancreas of alloxan induced diabetic mice

Article · January 2016

CITATIONS 2	5	reads 198				
3 autho	rs, including:					
	Madhuri Walvekar Shivaji University, Kolhapur 54 PUBLICATIONS 138 CITATIONS SEE PROFILE		Suvarna Pol Balwant College 8 PUBLICATIONS 20 CITATIONS SEE PROFILE			

All content following this page was uploaded by Madhuri Walvekar on 24 August 2016.

Research Article

Modulatory Effect of Fenugreek Loaded PLGA Nanoparticles on Lipofuscinogenesis in Pancreas of Alloxan Induced Diabetic Mice.

Walvekar M V*, Pol S B, Deshmukh V M

Department of Zoology, Shivaji University, Kolhapur

Available Online:31st December, 2015

ABSTRACT

Many traditional treatments have been recommended in the alterative system of medicine for the diabetes mellitus. Current research is now directed towards finding naturally occurring antidiabetic properties from plant origin. In Indian system of medicine Trigonella foenum graecum is an important medicinal plant and its leaves and seeds have been used in various ailments and as a health tonic. The purpose of this study was to examine the antioxidant activity of fenugreek seed extract (FSE) and its nanoparticles (FNPs) on body weight, pancreatic gland weight, lipid peroxidation and fluorescence product of the pancreas of alloxan induced diabetic mice. Adult albino male mice (Mus musculus L) were divided into four group's viz. i. Control Group: male mice were given subcutaneous injection of 0.15M acetate buffer pH 5.4 for 15 days. ii. Diabetic Group- mice were given single subcutaneous injection of alloxan 150 mg/kg body weight. iii. Diabetic \rightarrow FSE: mice were given subcutaneous injection of FSE at a dose of 15 mg/ kg body weight to diabetic mice for 15 days. iv. Diabetic \rightarrow FNPs: mice were given subcutaneous injection of FNPs at a dose of 15mg/ kg body weight to diabetic mice for 15 days. Body weight and pancreatic gland weight was reduced in diabetic group but increased in fenugreek supplementary group. The end product of lipid peroxidation malondialdehyde (MDA) and fluorescence product were increased in diabetic group and after fenugreek administration the level of both the parameters reduced significantly. The results suggest that an anti-lipid peroxidation activity of fenugreek seeds and its nanoparticles, nanoparticles treated group showed better results than fenugreek seed extract treated groups indicating that the fenugreek nanoparticles are best antidiabetic and antioxidant as compared fenugreek seed extract.

Key words: Fenugreek seed extract, fenugreek nanoparticles, lipid peroxidation, fluorescence product, diabetes mellitus.

INTRODUCTION

Diabetes mellitus (DM) is now a worldwide disease and in particular, the number of young patients is increasing^{1, 2}. The β cells in the pancreas are susceptible to ROS because they express very low levels of antioxidants³; therefore, it is considered that β cells are easily subjected to oxidative stress. It is well known that oxidative stress caused by ROS contributes to β cell death or dysfunction of the pancreas in type-1 diabetes⁴.

Alloxan is a toxic glucose analogue, which selectively destroys insulin producing cells in the pancreas when administered to rodents and many other animal species. This causes an insulin dependent diabetes mellitus (called alloxan Diabetes) in these animals, with characteristics similar to type-1 diabetes in humans⁵.

Management of DM may include lifestyle modifications, diet, exercise and long term use of hypoglycemic agents or insulin therapy⁶. It has been investigated that for a long time plants based herbal medicines or their extracts have been the major source of drugs for the treatment of DM in Indian medicine and other ancient systems, because plant products are frequently considered to be less toxic and more free from side effects than modern synthetic drugs⁸. Medicinal plants have become more important area of active research. Many herbs and plants have been

described as possessing hypoglycemic activity when taken orally^{7,9}. However, large floras are still waiting for investigation for their medicinal properties¹⁰. Medicinal plants possess antidiabetic potential or bioactive compounds such as glycosides, alkaloids, terpenoids, carotenoids, flavonoids and are confirmed to be effective in both preclinical and clinical studies^{11, 12}.

Fenugreek (*Trigonella foenum graecum*) is an annual herb that belongs to the family Leguminosae. The seeds of fenugreek are commonly used in India and in oriental countries as a spice in food preparations due to their strong flavor and aroma. The seeds are reported to have restorative and nutritive properties and to stimulate digestive processes¹³. Fenugreek seeds have been shown to have hypocholesterolemic effects in type 1 and 2 diabetes mellitus patients^{14, 15} and alloxan induced diabetic animals ^{16, 17}.

The hydrophobic character of fenugreek results in pharmacokinetic restrictions such as low absorption and bioavailability by oral route, extensive metabolism and rapid elimination¹⁸. Biodegradable polymeric nanoparticles are extensively used to improve the therapeutic properties of various drugs and bioactive compounds. The reasons for the widespread use of PLGA are its biodegradability, biocompatibility and the fact that

mice.	values are mea	III I v	S.D. (Numbers in parentin	esis denotes num		пппа	15).		
Sr.	Treatment		Weight of animal (gm)	Statistical	Weig	ght	of	pancreas	Statistical
No.	(n=5)			Significance	(mg)				Significance
1	Control		28.5296 ± 2.1605		193.8	875 :	± 15.	597	
2	Diabetic		22.2893 ± 2.5344	1:2, P<0.01	123.8	875 :	± 12.	3917	1:2, P< 0.01
3	Diabetic	\rightarrow	26.4085 ± 1.6625	2:3, P< 0.01	158.3	375 :	± 19.	9423	2:3, P< 0.01
	FSE			2:4, P<0.01					2:4, P<0.01
4	Diabetic	\rightarrow	27.2975 ± 0.9374	3:4, P-non-	185.7	$75 \pm$	9.80	89	3:4, P< 0.01
	FNPs			significant					

Table 1: Effect of fenugreek nanoparticles on body weight (gm) and weight of pancreas (mg) of alloxan induced diabetic mice. Values are mean \pm S.D. (Numbers in parenthesis denotes number of animals).

Table 2: Effect of fenugreek nanoparticles on the level of total lipid peroxidation (n mol MDA / mg wet tissue) and fluorescence product (μ g/ mg wet tissue) of alloxan induced diabetic mice. Values are mean \pm S.D. (Numbers in parenthesis denotes number of animals).

Sr.	Treatment	Total lipid peroxidation	Statistical	Fluorescence product	Statistical
No	(n=5)		Significance		Significance
1	Control	23.8163 ± 4.2049		0.004133±0.000945	
2	Diabetic	43.096 ± 2.9609	1:2, P<0.01	0.00917 ± 0.000859	1:2, P< 0.01
3	Diabetic \rightarrow FSE	28.7426 ± 4.7865	2:3, P<0.01	0.003921±0.000993	2:3, P< 0.01
4	Diabetic \rightarrow FNPs	23.3104 ± 3.9876	2:4, P<0.01 3:4, P-non- significant	0.003529±0.000856	2:4, P<0.01 3:4, P< 0.01

drug products containing PLGA have been approved for parenteral use by regulatory authorities around the world¹⁹. So to find out whether fenugreek nanoparticles will be more useful or FSE in diabetes we have synthesized PLGA nanoparticles. We have synthesized the fenugreek loaded PLGA nanoparticles, previously nobody has synthesized and we have reported 1st time these fenugreek nanoparticles.

MATERIAL AND METHODS

Preparation of fenugreek seed extract

Fenugreek seeds were collected from the Mahatma Phule Krushi Vidyapeeth, Rahuri. They were (10g) were cleaned and ground into a fine powder using a grinding machine. Ethanol was used for extraction by soxhelt extraction method²⁰. The extract was evaporated to dryness under reduced pressure at 60° C by rotary evaporator. Extract was placed in dark bottle and stored at -8°C.

Synthesis of fenugreek loaded PLGA nanoparticles (FNP's)

Fenugreek loaded PLGA based nanoparticles was prepared using oil in water single emulsion solvent evaporation process²¹.

Animals

Male albino mice (*Mus Musculus* L.) were used for present study. They were bred and reared in departmental animal house (1825/PO/EReBi/S/15/CPCSEA) in separate cages under proper conditions of light, temperature and humidity. They were supplied with Amrut mice feed (Pranav Agro industries) and water *ad libitum*.

Experimental design: Mice were divided in to four groups: 1) *Control Group*: Three months male mice were given subcutaneous injection of 0.15m acetate buffer pH 5.4 for 15 days.

2) *Diabetic Group:* Three months male mice were given subcutaneous injection of alloxan 150 mg /kg body weight for 15 days ²².

3) Diabetic \rightarrow FSE: Three months male mice were given subcutaneous injection of fenugreek seed extract at a dose of 15 mg /kg body weight to diabetic mice for 15 days ²³. 4) Diabetic \rightarrow FNPs: Three months male mice were given subcutaneous injection of fenugreek nanoparticles at a dose of 15mg/ kg body weight to diabetic mice daily for 15 days.

After completion of the dose, the mice were killed by cervical dislocation; pancreas were dissected out, blotted and weighed. The pancreas tissue was homogenized by using mixture containing 75 mM phosphate buffer (pH7.04), 1mM ascorbic acid, 1mM ferric chloride and 0.001 ml chlortetracycline (10ppm).

Measurement of body weight of mice

Mice were weighed before starting experiment, during respective treatment and also after completion of each treatment. The record of these observations was maintained.

Measurement of pancreatic gland weight of mice

The mice from respective groups were killed by cervical dislocation after completion of treatment. Pancreas dissected out, dried with the help of blotting paper and wet weight of gland was measured using digital scale balance. The record of these observations was maintained.

Determination of total lipid peroxidation ²⁴

Tissue homogenate was prepared in chilled mortar using 75mM potassium phosphate buffer pH 7.04 containing 1mM ascorbic acid and 1mM ferric chloride and the total lipid peroxidation was estimated.

Measurement of fluorescence product ²⁵

The lipofuscin granules from pancreas were extracted using chloroform: Methanol mixture (2:1 v/v). The fluorescence was measured by using quinine sulphate as a standard by using photofluorometer.

All values were expressed as mean \pm S.D. statistical analysis was carried out by one way ANOVA, Tukey's HSD test.

RESULTS

The average body weight and gland weight was 28 gm and 193 mg respectively at the beginning of the treatment. Diabetic mice showed a progressive reduction in body weight and gland weight as compared to control group. The body weight and pancreatic gland weight in FSE treated diabetic mice were significantly increased as compared to diabetic mice. (Table no. 1). The total lipid peroxidation and fluorescence product in pancreas was increased in diabetic group as compared to control and increase was significantly in Diabetic \rightarrow FSE and Diabetic \rightarrow FNPs (2:3, P < 0.01; 2:4, P < 0.01) as compared to diabetic group. The MDA and fluorescence product was decreased in Diabetic \rightarrow FNPs (2:3, P < 0.01; 2:4, P < 0.01) as compared to diabetic group. The MDA and fluorescence product was decreased in Diabetic \rightarrow FSE (Table 2).

DISCUSSION

In our study, we observed decrease in body weight, pancreas weight, while significant increase in fluorescence product and concentration of MDA, a secondary product of LPO, in pancreas of alloxan treated diabetic mice. Alloxan is a beta cytotoxin induces diabetes in a wide variety of animal species by damaging pancreatic B cells resulting in decrease in endogenous insulin release which lead to decrease glucose utilization by the tissues and a resultant diabetic (Hyperglycemia) condition.

Strain (1991) has observed disturbance of antioxidant defence system in DM i.e. oxidative stress play an important role in pathogenesis of diabetes²⁶. Oxidative stress is the result of excessive free radical production. These free radicals are scavenged by various antioxidants enzymes²⁷⁻²⁹. Unscavenged free radicals exert peroxidative effect on polyunsaturated fatty acids of the membrane of cells as well as cell organelles³⁰. Malondialdehyde is formed in extensive membrane lipid peroxidation. These peroxidized membranes are digested by lysosomes. The free radicals also bring about damage to lysosomes and lysosomal enzymes, making them inefficient which turn into residual bodies³¹⁻³³. These are lipofuscin granules. As lipofuscin granules are auto fluorescent they lead to increase in fluorescence product also³⁴. Lipofuscin is often called age pigment and considered a hallmark of aging. This is not only because the amount of lipofuscin increases with age, showing an almost linear dependence³⁵, but also and more importantly because the rate of lipofuscin accumulation correlates negatively with longevity³⁶⁻³⁹.

In the present study the level of lipid peroxidation and fluorescence product was decreased after treatment of fenugreek seed extract and fenugreek nanoparticles. These results suggest that FSE and FNPs administration in diabetic mice reduce LPO and MDA product possibly by decreasing free radical formation and increasing antioxidant. The administration of an antioxidant such as fenugreek seed extract may ameliorate tissue dysfunction since antioxidants are known to improve tissue integrity⁷. ^{40, 41}.

The reduction in the lipid peroxidation and fluorescence level in pancreas of mice after receiving fenugreek extract and its nanoparticles indicated that it is having free radical scavenging capacity which helps to prevent cellular damage and thereby reduce lipid peroxidation. It concludes that, fenugreek seeds are having antioxidant and anti peroxidative properties. The better results seen in fenugreek nanoparticles treated group than fenugreek seed extract treated groups, it clearly indicates that the fenugreek nanoparticles is best antidiabetic and antioxidant than fenugreek seed extract.

REFERENCES

- 1. Harjutsato V, Sjoberg L, Tuomilehto J. Time trends in the incidence of type1 diabetes in finish children: a cohort study. Lancet 2008; 371: 1777-1782.
- 2. Mayer Davis EJ, Beyer J, Bell RA, Dabelea D, Dagostino RJ, Imperatore G, Lawrence JM, Liese AD, Liu L, Marcovina S, Rodriguez B. Search for diabetes in youth study group: Diabetes in Africa American youth: Prevalence, incidence and clinical characteristics: the search for diabetes in youth study. Diabetes Care. 2009; 32 (Suppl 2): S112-S112.
- 3. Lenzen S, Dorinkgern J, Tiedge M. Low antioxidant enzyme gene expression in pancreatic islets compared with various other mouse organs. Free Radic Biol Med. 1996; 20:463-466.
- 4. Cnop M, Welish N, Jonas JC, Jorns A, Lenzen S, Eizirik DL. Mechanisms of pancreatic beta cell death in type 1 and type 2 diabetes: many differences, few similarities. Diabetes 2005; 54: S97-S107.
- 5. Lenzen S. The mechanism of alloxan and streptozotocin induced diabetes. Diabetologica. 2008; 51:216-226.
- Lawal HA, Atiku MK, Khelpai DG, Wannang NN. Hypoglycaemic and hypolipidaemic effect of aqueous leaf extract of *Murraya Koenigii* in normal and alloxan diabetic rats. Nigerian J Physiol Sci. 2008; 23(1-2): 37-40.
- Akhtar FM, Ali MR. Study of antidiabetic effect of a compound medicinal plant prescription in normal and diabetic rabbits. Journal of the Pakistan medical Association 1984; 34: 239-244.
- Brinker FJ. Herb Contraindications and drug interactions: With Appendices Addressing specific conditions and medicines. 2nd. Edn., Eclectic Medical Publications, Oregon, USA., 1998; ISBN:9781888483062, Pages:263.
- Pepato MT, Baviera AM, Vendramini RC, Perez MPMS, Kettelhut IC, Brunetti IL. Cissus Sicyoides (Princess Vine) in the long term treatment of streptozotocin diabetic rats. Biotechnol Appl Biochem. 2003; 37:15-20.
- Kirtikar KR, Basu BD. Indian Medicinal Plants. Vol. 3.
 2nd ed. In: Kirtikar KR, Basu BD (eds). Dehra Dun, India: International book distributors, 1987; 2061-2062.
- 11. Loew D, Kaszkin M. Approaching the problem of bioequivalence of herbal medicinal products. Phytother Res. 2002; 16:705-711.

- 12. Marles RJ, Farnsworth NR. Antidiabetic plants and their active Constituents. Phytomedicine. 1995; 2: 133-189.
- 13. Khosla P, Gupta DD, Nagpal RK. Effect of *Trigonella foenum graecum* (Fenugreek) on serum lipids in normal and Diabetic rats. International Journal of Pharmacology 1995; 27:89-93.
- 14. Gupta A, Gupta R, Lal B. Effect of *Trigonella foenum* graecum (fenugreek) seeds on glycaemic control and insulin resistance in type-2 diabetes mellitus: a double blind placebo controlled study. J. Assoc physicians India 2001; 49: 1057-1061.
- 15. Madar Z, Abel R, Samish S, Arad J. Glucose- lowering effect of fenugreekin non- insulin dependent diabetics . Eur J. Clin. Nutr. 1998; 42 (1):51-54.
- 16. Jelodar GA, Maleki M, Motadayen MH, Sirus S. Effect of fenugreek onion and garlic on blood glucose and histopathology of pancreas of alloxan induced diabetic rats. Indian J. Med Sci 2005; 59(2): 64-69.
- 17. Raju J, Gupta D, Rao AR, Yadava PK, Baquer NZ. *Trigonella foenum graecum* (fenugreek) seed powder improves glucose homeostasis in alloxan diabetic rat tissues by reversing the altered glycolytic, gluconeogenic and lipogenic enzymes. Mol. Cell. Biochem. 2001; 224(1-2): 45-51.
- Sharma RD, Raghuram TC, Rao NS. Effect of fenugreek seeds on blood glucose and serum lipids in type I diabetes. Eur J Clin Nutr 1990; 44: 310-306.
- 19. Fredenberg S, Wahlgren M, Reslow M. Axelsson A. The mechanism of drug release in poly (lactic-coglycolic) acid- based drug delivery systems: A review. Int J Pharm 2011; 415(12): 34-52.
- 20. Lim SN, Cheung PCK, Ooi VEC, Ang PO. Evaluation of antioxidative activity of extracts from a brown seaweed, *Sargassum siliquastrum*, J. Agri Food Chem. 2002; 50: 3862-3866.
- 21. Jaiswal J, Gupta SK, Kretuter J. Preparation of biodegradable cyclosporine nanoparticles by high pressure emulsification solvent evaporation process. J Control Release 2004; 96: 169-178.
- 22. Al-Shamaony L, Al-Khazraji SM, Twaiji HA. Hypoglycemic effect of *Artemisia herba alba* II. Effect of a valuable extract on some blood parameters in diabetic animals. J. Etanopharmacol 1994; 10: 167-171.
- 23. Vijayakumar MV, Manoj KB. Hypoglycemic effect of a novel dialysed fenugreek seeds extract is sustainable and is medicated, in part, by the activation of hepatic enzymes. J. Phytother. Res. 2008; 22: 500-505.
- 24. Wills ED. Mechanisms of lipid peroxide formation in animal tissue. J. Biochem. 1966; 99: 667-676.
- 25. Dillard CJ, Tappel AL. Fluorescent products of lipid peroxidation of mitochondria and microsomes. Lipids 1971; 6: 715-721.
- 26. Strain J. Disturbances of micronutrient and antioxidant status in diabetes. Proc Nutr Soc. 1991; 50:591-604.

- De Marchena O, Gumieri M, Mekharn G. Glutathione peroxidase levels in brain. J Neurochem. 1994; 22: 773-776.
- Chance B, Sies H, Boveris A. Hydroperoxide metabolism in mammalian organs. Physiology. 1979; 59:527-605.
- 29. Forman HJ, Kennedy J. Dihydrolate dependent superoxide production in rat brain and liver. A function of primary dehydrogenase. Arch Biochem Biophys. 1976; 173: 219-224.
- 30. Schroeder F. Role of membrane lipid asymmetry in aging. Neurobiol Aging. 1984; 5: 323-333.
- 31. Ivy GO, Schottler F, Wenzel J, Boudry M, Lynch G. Inhibitors of lysosomal enzymes: Accumulation of lipofuscin like dense bodies in the brain. Science. 1984; 226: 985-987.
- 32.Zs Nagy, I: An attempt to answer the questions of theoretical gerontology on the basis of membrane hypothesis of aging. In Advances in Biosciences, London, Pergamon, vol. 64, 1987: 393-413.
- 33. Thakkar BK, Dastur DK, Manghani DK. Neuropathology and pathogenesis of experimental fenfluramine toxicity in young rodents. Indian J Med Res. 1990; 13(92) 54-65.
- 34. Walvekar MV, Sarvalkar PP, Pol SB. Effect of curcumin supplementation on the submandibular salivary glands of D-galactose induced male mice. IJPSR. 2012; 3(12): 4796-4800.
- 35. Strehler BL, Mark DD, Mildvan AS, Gee MV. Rate of magnitude of age pigment accumulation in the human myocardium. J Gerontol. 1959; 14: 430–439.
- Munnel JF, Getty R. Rate of accumulation of cardiac lipofuscin in the aging canine. J Gerontol 1968; 23:154–158.
- Nakano M, GotohS . Accumulation of cardiac lipofuscin depends on metabolic rate of mammals. J. Gerontol. 1992; 47:B126–B129.
- 38. Hosokawa H, Ishii N, Ishida H, Ichimori K, Nakazawa H, Suzuki K. Rapid accumulation of fluorescent material with aging in an oxygen-sensitive mutant mev-1 of *Caenorhabditis elegans*. Mech Ageing Dev. 1994; 74:161–170.
- 39. Sheehy MR, Greenwood JG, Fielder DR. Lipofuscin as a record of "rate of living" in an aquatic poikilotherm. J Gerontol.1995; 50: B327–B336.
- 40. Deshmukh VM, Walvekar MV, Desai SR. Modulatory effect of fenugreek (*Trigonella foenum graecum*) seed extract on salivary glands lipofuschinogenesis in aging accelerated male mice. Int J Pharm Bio Sci 2014; 5 (2): (B) 83 – 89.
- 41. Walvekar MV, Deshmukh VM, Pol SB. Structural and Cytochemical Study of Salivary Glands after Fenugreek Seed Extract Administration in Oxidatively Stressed Mice. International Journal of Pharmacognosy and Phytochemical Research 2015; 7(3): 395-400.