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## EFFECT OF CURCUMIN SUPPLEMENTATION ON THE SUBMANDIBULAR SALIVARY GLANDS OF D-GALACTOSE INDUCED MALE MICE

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### Keywords:

Aging, Curcumin, Lipid peroxidation, Submandibular gland, Lipofuscin granules

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### ABSTRACT

Significant increase in the salivary gland lipid peroxidation, fluorescence product as well as accumulation of lipofuscin granules–age pigment was observed after injection of 5% D-galactose 0.5 ml daily for 30 days. Supplementations of turmeric extract (30mg/kg body weight) for 30 days during and after D-galactose treatment significantly reduced all the above parameters. Histochemical changes in the form of accumulation of age pigment were observed in the submandibular gland (SMG) of experimental animals. The normalization of levels of all the studied parameters in the SMG by curcumin supplementation in D-galactose induced aging accelerated mice is suggestive of the possible ameliorating effect of curcumin on SMG. While in curative group II the levels of all parameters were elevated as compared to D-galactose treated group mice.

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**INTRODUCTION:** Reactive Oxygen Species (ROS) are major sources of primary catalyst produced in our body during aerobic metabolism that initiates oxidation *in vivo* and *in vitro* and are created oxidative stress which results in numerous diseases and disorders such as cancer<sup>1</sup>, Alzheimer disease<sup>2</sup>, aging<sup>3</sup>.

Organisms have evolved a wide array of enzymatic and nonenzymatic endogenous antioxidant defenses to balance the physiological generation of free radicals. But in situation of increased free radical generation the reinforcement of endogenous antioxidants with dietary antioxidants may be particularly important in diminishing the cumulative effects of antioxidants of oxidatively damaged molecules. Antioxidants are mainly involved in neutralization of oxidants. Natural antioxidants are the secondary metabolites of phytochemicals<sup>4</sup>.

Lipofuscin pigments commonly referred as age pigments since the histological and biochemical observations that pigments concentration in cells increases with age<sup>5-10</sup> are proposed to be the stable metabolic end products of *in vivo* lipid peroxidation<sup>11-12</sup>.

Medicinal herbs that are relatively safe, cheap and easily available constitutes the cornerstone of traditional medicinal practice worldwide. These medicinal plants represent a great deal of untapped reservoir of the drugs and the structural diversity of their component molecules makes a valuable source of novel lead compounds<sup>13</sup>.

*Curcuma longa* (Family - Zingiberaceae) is a perennial shrub commonly called as 'turmeric'. Curcumin, a yellow pigment in turmeric, has received attention as a promising dietary supplementation for the protection against fibrogenic insults<sup>14,15</sup>.

A wide range of biological and pharmacological activities of curcumin have been investigated by Okada<sup>16</sup>. Hepatoprotective, oxidative, antimutagenic and anticarcinogenic effect of curcumin have been shown by previous studies of Premkumar<sup>17</sup>.

D-galactose is a normal reducing sugar in the body. At its normal level, it is usually converted into glucose by galactose-1-phosphate uridyltransferase and galactokinase. However, at high levels, it can be oxidized into aldehydes and H<sub>2</sub>O<sub>2</sub> in presence of galactose oxidase<sup>18</sup>. Administration of D-galactose induces symptoms similar to those of normal aging; many investigations using rodents have demonstrated that the injection of D-galactose can lead to excessive formation of ROS, neural damage and a significant decline in learning and memory capacity<sup>19-20</sup>.

**MATERIAL AND METHODS:** 30 Swiss albino male mice *Mus musculus* weighing 50-55 gm were used for present study. The mice were kept in clean and dry plastic cages having bedding of rice husk and with 12 hr light-dark cycle at standard environmental conditions. The animals were fed with palletized commercial mice chow (Amrut mice feed, Sangli, MS, India), without any restriction to food or drinking water during the experimental period.

Mice were assigned into 5 groups of six animals each. The control group mice received 0.5 ml of sterile distilled water (sc) daily for 30 days. Animals in aging accelerated group were injected with 0.5 ml of 5% D – galactose /day for 30 days. Mice in protective group receive D-galactose (sc) + curcumin (obtained from Sigma, USA- 30mg/kg body weight/day) orally for 30 days. In curative groups, I and II, animals receive D-galactose (sc) for 30 days and then were orally fed with once daily with curcumin for next 30 and 45 days respectively.

All animals were treated in accordance with the CPCSEA. After 24 hr of completion of treatment, all animals were sacrificed by cervical dislocation, SMG were removed, weighed, freezeed and homogenized in respective homogenization media.

1. Total and mitochondrial lipid peroxidation was done by Wills<sup>21</sup> method.
2. Fluorescence was measured by Dillard and Tappel<sup>12</sup> method.
3. Histochemical demonstration of lipofuscin granules was done by Zeihl Nelson Carbol fuscine method<sup>22</sup>.

For histochemical demonstration of lipofuscin granules, SMGs were fixed in NBF, hydrated to remove excess fixative and then dehydrated through alcohol grades, cleared in xylene and embedded in paraffin wax. 5µm thick sections were taken with the help of microtome (Yorco, Lipshaw type).

## RESULT:

- A. Assay of lipid peroxidation and fluorescence product:** The MDA content as well as fluorescence product of submandibular gland was increased significantly in D-galactose treated group, as compared to control. While in protective and curative group I the same parameters were decreased as compared to D-galactose treated group. And the decrease was significant as compared to D-galactose induced aging accelerated group in each group. In case of curative group II the increase in MDA content as well as fluorescence as compared to D-galactose treated group was observed (**Table 1 and 2**).

**TABLE 1: LIPID PEROXIDATION (N MOLES MDA /MG WET WEIGHT OF TISSUE) IN SMG OF AGING INDUCED MALE MICE AND EFFECT OF CURCUMIN ON THE SAME** (Values are mean ± S.D.)

S/N	Groups	Age of animals (in weeks)	Total lipid peroxidation	Statistical significance	Mitochondrial Lipid peroxidation	Statistical significance
1	Control (6)	24	11.589±0.074		20.83±0.01581	
2	Aging Accelerated (6)	28	34.648±0.0031	1:2 P<0.001	69.2944±0.05292	1:2P<0.001
3	Protective (6)	28	17.362± 0.0530	2:3 P<0.001	57.6518±0.003651	2:3 P<0.001
4	Curative I (6)	32	11.57±0.0484	2:4 P<0.001	23.0773±0.0274	2:4 P<0.001
5	Curative II (6)	34	51.9056±0.0118	2:5 P<0.001	103.832±0.03564	2:5 P< 0.001

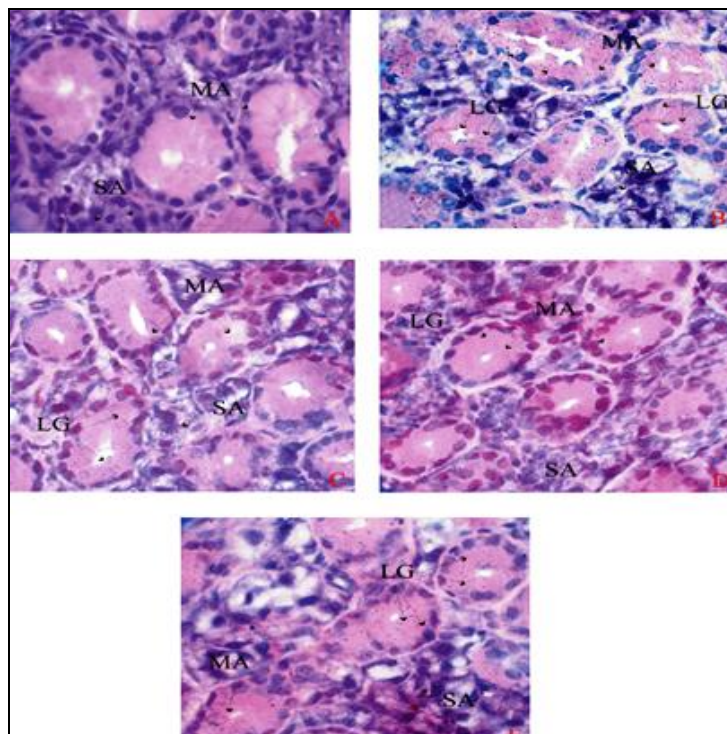
(Numbers in parenthesis denoted number of animals)

**TABLE 2. FLUORESCENCE PRODUCT IN THE SMG OF AGING INDUCED MALE MICE AND EFFECT OF CURCUMIN FEEDING ON THE SAME**  
(Values are mean  $\pm$  S.D.)

S/N	Groups	Age of animals (in weeks)	Fluorescence product	Statistical significance
1	Control (6)	24	0.00184 $\pm$ 0.000089	
2	Aging Accelerated (6)	28	0.00585 $\pm$ 0.000084	1:2 P<0.001
3	Protective (6)	28	0.0019093 $\pm$ 0.000011	2:3 P<0.001
4	Curative I (6)	32	0.001886 $\pm$ 0.00002	2:4 P<0.001
5	Curative II (6)	34	0.005420 $\pm$ 0.00022	2:5 P< 0.001

(Numbers in parenthesis denoted number of animals).

**B. Lipofuscin granules:** The light microscopic observation of lipofuscin granules in SMG was displayed in figure. The increased accumulation of lipofuscin granules were observed in SMG of aging accelerated group, as compared to control group (**Fig. 1A, B**). While in curcumin co-treated group, (Protective) there was decrease in accumulation of lipofuscin granules as compared to aging accelerated group (**Fig. 1C**). A similar type of changes was observed in curative I group (**Fig. 1D**). But in case of curative II group, number of lipofuscin granules was increased as compared to curative I group (**Fig. 1E**).



**FIG. 1A: CROSS SECTION OF SUBMANDIBULAR GLAND OF CONTROL GROUP MICE.**

**FIG. 1B: CROSS SECTION OF SUBMANDIBULAR GLAND OF D-GALACTOSE TREATED GROUP MICE.**

**FIG. 1C: CROSS SECTION OF SUBMANDIBULAR GLAND OF PROTECTIVE GROUP MICE.**

**FIG. 1D: CROSS SECTION OF SUBMANDIBULAR GLAND OF CURATIVE GROUP I MICE.**

**FIG. 1E: CROSS SECTION OF SUBMANDIBULAR GLAND OF CURATIVE GROUP II MICE.**

**MA: MUCOUS ACINAR CELLS, SA: SEROUS ACINAR CELLS, LG : LIPOFUSCIN GRANULES.**

**DISCUSSION:** Normal biochemical processes produce free radicals in the cells. Thus the cells are always subjected to the attack of free radicals<sup>23</sup>. These radicals are scavenged by various antioxidant enzymes such as glutathione peroxidase<sup>24</sup>, catalase<sup>25</sup> and superoxide dismutase<sup>26</sup>. Unscavenged free radicals exert peroxidative effect on polyunsaturated fatty acids of the membrane of cells as well as cell organelles<sup>27</sup>.

Malonicdialdehyde is formed in extensive membrane lipid peroxidation. These peroxidized membranes are digested by lysosomes. The free radicals also bring about damage to lysosomes and lysosomal enzymes, making them inefficient which turn into residual bodies<sup>28-30</sup>. These are the lipofuscin granules. As lipofuscin granules are auto fluorescent they lead to increase in fluorescence product also.

The accumulation of lipofuscin pigment as a function of age, oxidative stress and antioxidant deficiency has been known for many years. Present knowledge suggests that these pigments results from lipid peroxidation<sup>31</sup>.

Lipofuscin is often called age pigment and considered a hallmark of aging. This is not only because the amount of lipofuscin increases with age, showing an almost linear dependence<sup>32</sup>, but also and more importantly because the rate of lipofuscin accumulation correlates negatively with longevity (i.e. positively correlates with the rate of aging)<sup>33-36</sup>.

It has been shown repeatedly that oxidative stress promotes lipofuscin formation, whereas antioxidant defense prevents it.

Accelerated lipofuscinogenesis was observed in fibroblast, glial cells, cardiac myocytes and retinal pigment epithelium (RPE) cells with cultured at high (40%) ambient oxygen<sup>31</sup>.

Decrease in lipofuscin pigment after curcumin feeding curative group- I could be explained as a result of cell proliferation. Conceivably, proliferating cells may replace older lipofuscin-loaded ones. Curcumin has decreased the lipofuscin granules in the sub-mandibular glands and thereby resulted in highly significant decrease in fluorescence. While in curative group II in which mice were fed with curcumin for 45 days increase in lipofuscin pigments were observed. This may be due to long term administration of curcumin leads to accumulation of lipofuscin granules.

The antioxidant mechanism of curcumin has recently been a focus of interest of free radical chemist and biologist<sup>37-40</sup>. Barclay and collaborators compared the antioxidant activities of curcumin and dimethyl-curcumin against free radical initiated peroxidation of styrene in chlorobenzene solution and concluded that curcumin is a classical phenolic chain breaking antioxidant, donating H –atoms from the phenolic groups<sup>37</sup>. Recently, Priyadarshani<sup>39</sup> also compared the antioxidant activity of curcumin and dimethylcurcumin against radiation-induced lipid peroxidation of rat liver microsomes and the free radical-scavenging activity against 2, 2- diphenyl-1 picryltry-drazyl (DPDH) and concluded that the phenolic group plays a major role in the activity of curcumin.

**CONCLUSION:** In conclusion, curcumin is effective antioxidant which can protect mice SMG cells from free radical induced oxidative stress and the H-atom abstraction from the phenolic group is responsible for its anti-oxidative property. Its lengthy history as a foodstuff might lead some consumers to conclude that turmeric is completely safe to use as a health aid. However, like virtually every other medicinal substance, man-made or natural, turmeric is not free from side effects, especially when taken in excessive doses.

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